

REMARKS

Reconsideration is respectfully requested.

Claims 1-50, 66, 70, 73, and 89 have been cancelled. Claims 51-65, 67-69, 71, 72, 74-88, and 90-93 are pending. Applicants also have filed a request for continued examination and supplemental information disclosure statement with this response.

With respect to all amendments and cancelled claims, Applicant has not dedicated or abandoned any unclaimed subject matter and moreover have not acquiesced to any rejections and/or objections made by the Patent Office. Applicant reserves the right to pursue prosecution of any presently excluded claim embodiments in future continuation and/or divisional applications.

Withdrawn Rejections and Objections

Applicants respectfully thank the Examiner for withdrawing the rejections and objections in the office action dated December 12, 2005.

Declaration Under 37 C.F.R. § 1.131

The Examiner objected to the declaration under 37 C.F.R. § 1.131 previously submitted on July 14, 2004 because the declaration was unsigned. Applicants herewith resubmitted a signed declaration in accordance with 37 C.F.R. § 1.131, as discussed below.

Information Disclosure Statement.

The Examiner states that reference C10 was not submitted to the Office.

Applicants herewith include a Supplemental Information Disclosure Statement including reference C10. Applicants respectfully request its consideration.

Claim Interpretation

The Examiner states that “the term ‘electrode’ will be considered as meaning ‘a solid support comprising a metallic surface.’”

The Examiner also states that “the term ‘shielding’ is used here in its everyday meaning, i.e. ‘preventing physical contact.’ Therefore, ‘blocking moieties shielding nucleic acids from the electrode’ means any structural elements which prevent contact target and/or probe nucleic acids with the electrode.”

Applicants respectfully traverse these interpretations. Applicants do not take a position with respect to the interpretation, as no such position is necessary given the accompanying declaration.

35 USC § 102(e)

Claims 51-58, 60-62, 64-73, 79, 80, 82, 83, 85-89 and 93 stand rejected under 35 USC § 102(e) as anticipated by Wohlstadter et al., US Patent No. 6,066,448 (“Wohlstadter”). Wohlstadter is a continuation-in-part of USSN 08/402,076, filed March 10, 1995, which is a continuation-in-part of USSN 08/402,277, also filed March 10, 1995. Accordingly, the earliest possible priority date available for the disclosure in Wohlstadter is March 10, 1995.

The Applicants herewith submit a Declaration under 37 C.F.R. §1.131 by inventors Thomas J. Meade and Jon F. Kayyem. The Declaration demonstrates that the claimed invention was made prior to the earliest possible March 10, 1995 priority date of Wohlstadter.

To antedate a 35 USC 102(e) reference, the inventors must show possession of a species within a claimed genus. As stated in the MPEP:

[t]he 37 CFR 1.131 affidavit or declaration must establish possession of either the whole invention claimed or something falling within the claim (such as a species of a claimed genus), in the sense that the claim as a whole reads on it. *In re Tanczyn*, 347 F.2d 830, 146 USPQ 298 (CCPA 1965). MPEP 715.02.

The inventors need not show possession subject matter identical to that of the references. As further stated in the MPEP:

a 37 CFR 1.131 affidavit is not insufficient merely because it does not show the identical disclosure of the reference(s) or the identical subject matter involved in the activity relied upon. If the affidavit contains facts showing a completion of the invention commensurate with the extent of the invention as claimed is shown in the reference or activity, the affidavit or declaration is sufficient, whether or not it is a

showing of the identical disclosure of the reference or the identical subject matter involved in the activity. See *In re Wakefield*, 422 F.2d 897, 164 USPQ 636 (CCPA 1970). MPEP 715.02.

The Declaration and associated Exhibits demonstrate that the claimed invention was made prior to the earliest Wohlstadter priority date. Paragraph 5 of the Declaration summarizes an embodiment of the invention within the scope of claim 51.

The Declaration discloses the production of an array of claim 51 depicted in Exhibit A. Different regions on the array are defined by 8x8 micron squares on the photolithographic mask. The gold surface is the electrode of claim 51. The thiol-(CH₂)₁₆-OH is the blocking and linking moiety. When the thiol-(CH₂)₁₆-OH is covalently attached to the nucleic acid and the gold surface, it is the linking entity. When the thiol-(CH₂)₁₆-OH is attached to the gold surface, and not attached to the nucleic acid, it is the blocking moiety.

The fluorescent complement is an agent that distinguishes between single stranded and double stranded nucleic acids. Dark squares indicate locations where single stranded nucleic acids were ablated off, and light squares indicate where nucleic acid hybrids were present. A montage of images is depicted in Exhibit A.

The declaration outlines that the invention was completed in this country prior to March 10, 1995. Because the claimed invention was made prior to the earliest Wohlstadter priority date, the Wohlstadter reference is not prior art. Applicants respectfully request withdrawal of this ground for rejection.

35 USC § 103(a)

Claims 59, 63, 81, 84 and 85 stand rejected under 35 USC § 103(a) as being unpatentable over Wohlstadter in view of Kayyem et al., U.S. Patent No. 6,096,273 (Kayyem).

In a separate rejection, claims 75-78 and 90-92 also stand rejected 35 USC § 103(a) as being unpatentable over Wohlstadter in view of Kayyem.

As demonstrated above in the response to the rejection under 35 USC § 102(e), Wohlstadter is not a prior art reference. Therefore, Wohlstadter cannot be combined with Kayyem as described by the Examiner in satisfaction of the requirements of 35 USC § 103(a). Accordingly, Applicants respectfully request withdrawal of this ground for rejection.

CONCLUSION

On the basis of the amendments and remarks presented herein, Applicants believe that this application is in a condition of allowance. Applicants respectfully request that the Examiner pass this application to issue, and early notification of such is requested.

If the Examiner has any questions, she is invited to call the undersigned at (415) 781-1989.

Respectfully submitted,
DORSEY & WHITNEY LLP

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Filed Under 37 C.F.R. § 1.34



PATENT

Attorney Docket No.: A-64411-2

Attorney File No.: 468267-00067

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of

MEADE *et al.*

Serial No.: 09/921,645

Filed: August 3, 2001

For: *Metallic Solid Supports Modified with Nucleic Acids*

Examiner: STRZELECKA, Teresa, E.

Group No. 1637

“EXPRESS MAIL” LABEL NO.:
EV 554099109 US

DECLARATION PURSUANT TO 37 C.F.R. § 1.131

Mail Stop RCE
Commissioner for Patents
P.O. Box 1450
Alexandria, VA 22313

Sir:

We, Thomas J. Meade and Jon F. Kayyem hereby declare as follows:

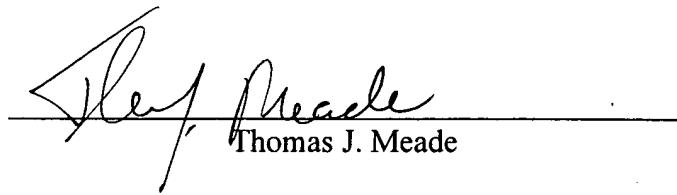
1. We are the inventors on the above-identified patent application and are familiar with its contents. We have also reviewed the pending claims in this application.
2. We are familiar with the Office Action mailed on November 17, 2003 wherein claims 51-69, 71, 72, and 74-93 were rejected over Wohlstadter et al. (6,066,448) which has an earliest possible priority date of March 10, 1995.
3. All of the ideas detailed in the above-identified application were contemplated in this country prior to March 10, 1995. This is evidenced by the appended documents.
4. One of the goals of the project that led to the filing of the parent application was to create a surface comprising a self-assembled monolayer with single stranded nucleic acids attached (referred herein as probes), and then to answer three questions: first, whether a solution-based complementary strand would bind to the probe; second, would a complementary strand attached to an atomic force

microscopy (AFM) tip bind to the probe, and if so; third, whether or not we could determin the force necessary to “tear apart” the duplex.

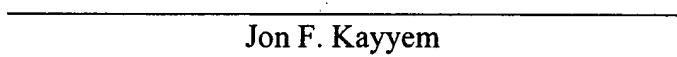
5. The experiments started out with the synthesis of the monolayer portion using an HO-(CH₂)₁₆-OH to form a molecule with a protected sulfur group (for attachment to a gold surface) on one end, to which a phosphoramidited nucleic acid was attached. The experiments proceeded with the coating of a gold surface with this monolayer-forming material. A photolithographic mask, with 8 x 8 micron squares on it, was then used to cover the gold surface. The surface was then exposed to a photoactivated agent and a mercury arc lamp which resulted in the ablation nucleic acids from the squares not covered by the mask. We then added a fluorescent complement to the surface, and viewed it under a confocal microscope. This resulted in a pattern of “light”, e.g. fluorescent, background, where the fluorescent solution based probes were found, and “dark” squares, where the surface-bound single stranded nucleic acid had been ablated off, and therefore no fluorescent probe was detected. A montage of several of these images, made over the course of the experiments, is attached as Exhibit A.
6. With regard to timing of these experiments, the documents attached as Exhibit B are pages from the notebook detailing the synthesis of some of the compounds used in these experiments. (Please note that all experiments not relevant to the present discussion have been redacted, as have all dates.) For example, page 136 documents the conversion of the HO-(CH₂)₁₆-OH molecule to the asymmetrical HO-(CH₂)₁₆-OAc needed for further reactions. The bottom of page 139 and the top of page 140 show the synthesis of the protected thiol-(CH₂)₁₆-OH molecule. the top of page 141 shows the reaction of the protected thiol-(CH₂)₁₆-OH molecule added to a phosphoramidite moiety. In conclusion, the invention was completed in this country prior to March 10, 1995.
7. We declare further that all statements made herein of our own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that the making of willful false statements and the like are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such

willful statements may jeopardize the validity of the application or any patent issuing thereon.

Date: 5-4-05


Thomas J. Meade

Date: _____


Jon F. Kayyem

willful statements may jeopardize the validity of the application or any patent issuing thereon.

Date: _____

Thomas J. Meade

Date: May 18, 2005


Jon F. Kayyem

EXHIBIT A

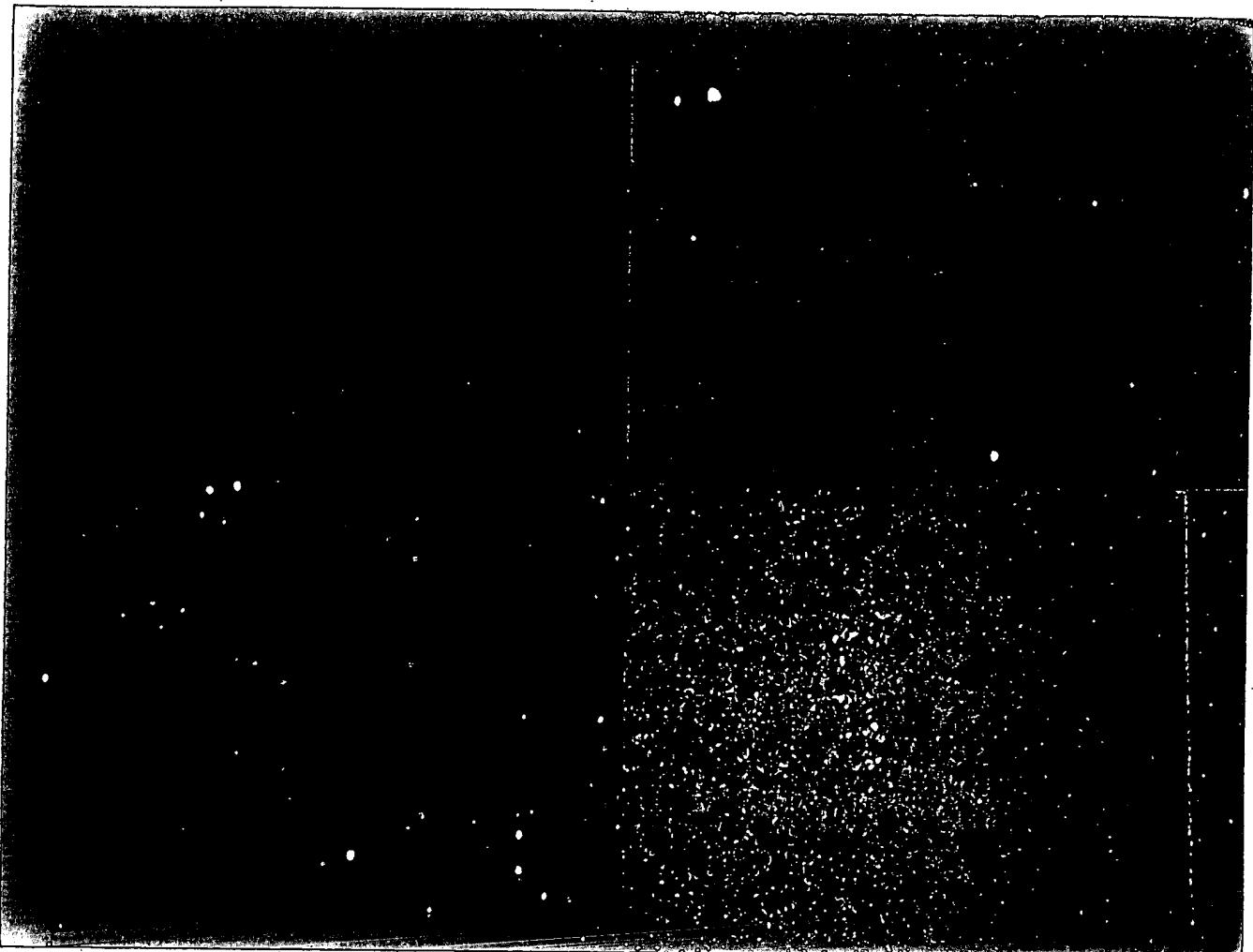
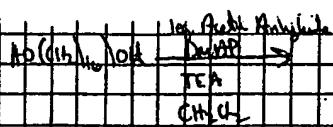


EXHIBIT B



0.5 g of $\text{Hg}(\text{C}_6\text{H}_5)_2\text{OAc}$ (mw 258.45; 1.9×10^{-3} molar) was placed in a small round bottom and 20 ml of CH_3Cl added along with 0.05 equiv (9.7×10^{-5} molar) or 11.8 mg) and 1.4 equiv of TEA (4.7×10^{-4} molar) and 1 equiv of Acetic Anhydride (mw 102.09; $d = 1.08$) (10.57 mg (1.08) or 173 μl).



25.50 atm/kmol

100%
std. press

constant volume

10/10
other / hexane

Refuge 2

2.05 g (7.93×10^{-3} molar) was placed in a 100 ml RBF and 60 ml s of CH_3Cl added along with 0.05 equiv DMSO (48 mg) and 1.4 equiv of TEA (17 ml) and 6.95 ml of Acetic Anhydride (1.08) and 100 ml of

137

138:

Run B

0.1 (dm₃), 0.1 + Acetone, DMSO, DMSO
TEA
DMSO

1.37 ml of Diol were dissolved in 75 ml of DMSO/0.1 M TEA

2 g DMAP (32.33 mg) were added and 1.4 equiv of TEA (1.4 ml's)

and 475 μ l of Pd-BaMgB₁₂

Tube 71

new
needle

50 ml solvent form 1 ml diol 0.1

12 mm needle

30 mm needle = 1 ml 0.1

The first column was

200 ml's of 9:1 hex:

1.2 liters of 180:20 hex/EtOH

was passed through and the

gradient pushed up to 30

55:45, after tube 72

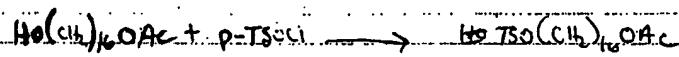
it took about 20 ml's of 55:45

to remove all the gradient

CH₃NO

500.93

Li⁺Mg²⁺



139

Finest. Form. page 1180

500mg (1.7 x 10⁻³ mols) of HO(CH₂)₁₆COCl₂ in in dry pyridine
and in 5 ml ethyl acetate and cooled to 0°C. & TSCl
(190.65) in 1 ml excess (or 634 mg) with a stirring bar
and allowed to proceed for 40 hrs. The solution was poured
into a beaker with 200 ml of ice water and suction filtered
at this stage. The filtrate was
stirred in heptane (20) and returned to

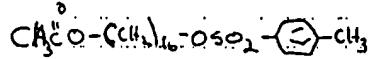
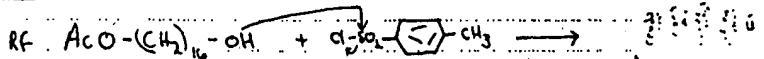
RF
Solvent front
48 mm
Rf = 12 mm = .25
Tos = 28 mm = .58 product
50:50
water:Et₂O
DIB →



TSCl
The product spot is UV active
and again inactive
probably the hydroxyl in the
C₁₆ chain displaces the acyl
on TSCl giving the
inactive (T) isomer.

degree. H-NMR is consistent with the proposed structure

Repeat



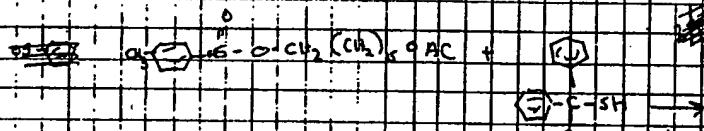
Solvent
Front = 47
Tos 215
Rf: 0.59

Tos: Rf = .36 MW ~~25.105~~ = 454.72

C₂₅H₄₂O₅S = 438.72
+ 15.958

0.0

140



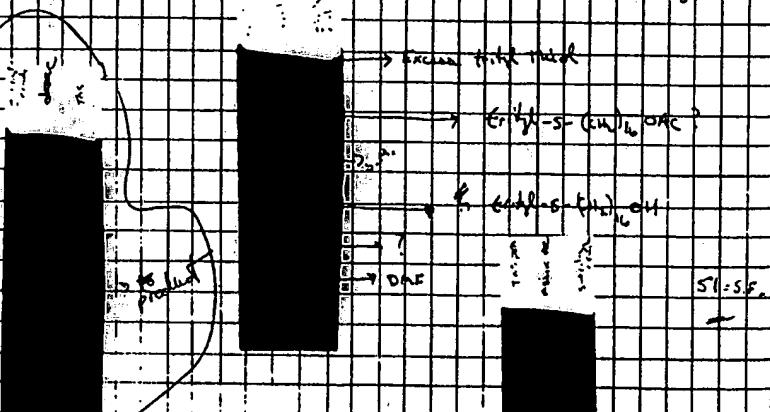
370 mg of $\text{TSO}(\text{CH}_3)_6\text{OAC}$ was dissolved in 10 ml of DNF and thoroughly degassed on the vac. line
($\mu\text{mol} = 45.72$ or 8.6×10^{-3} μmole)

$\text{NaOEt} \cdot 4\text{H}_2\text{O}$

- Triphenyl methyl boronate ($\text{MW} = 276.50$; $0.95 \text{ equiv} = 7.7 \times 10^{-3}$ μmole
 $= 213 \text{ mg}$)
- 38.8 mg of NaOEt (0.98 equiv) in 180 ml of H_2O

5 ml of ethanol was degassed on the vac. line and 213 mg of triphenyl boron added. 180 ml of degassed NaOEt in H_2O was added via syringe under N_2 . To this solution 370 mg of $\text{TSO}(\text{CH}_3)_6\text{OAC}$ in DME/EtOH was added and the solution degassed.

After treatment with NaOEt in MeOH



$\text{S}^1 = \text{S}^2$

solvent: $4\text{H}_2\text{O}$
 $\text{EtOH} = 1\text{ ml}$
 $\text{or } 8.6 \text{ ml}$

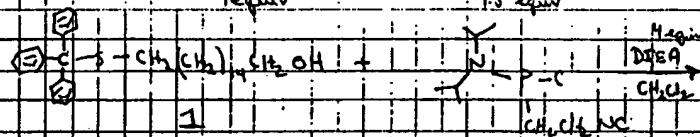
EtOH

The material was taken up in MeOH/MeOH 6:1

equiv

1.5 equiv

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• 220 mg of 1 = 4.4×10^{-3} molar

• 6.5×10^{-3} molar of phosphoroanhydrite $236.68 \text{ g/mol} \times 1/1061 \text{ M} = 4.48 \text{ molar}$

5.7 molar $\times 2.0 \times 10^{-3} \times 1.7 \times 10^{-3} \text{ molar } \text{DIEA} \text{ mw } 119/127.25 \frac{0.792}{127.25} = 5.7 \text{ molar}$

220 mg of 1 was slurried in 15 ml of dry CH_2Cl_2 and immediately 326 μl of DIEA was added. 145 μl of cyanethyl phosphorus(III) were added dropwise. After 30 min and addition of 50 μl of cyanethyl reagent was added.

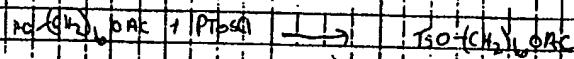
Note:

TEA MUST be present during flush!

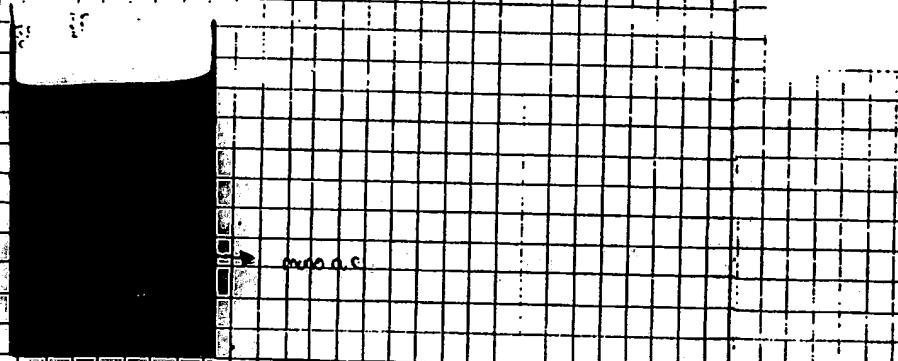
∴ At the addition of P^{III}
1% TEA to the reagent
phine, only the chloro
ether

Ti^{2+} O^{2-}

142



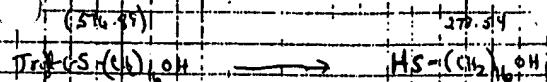
57 g of monac product were slurried in 300 ml. of dry pyridine and cooled to 0°C. 650 mg of TsCl was added and the reaction mixture allowed to proceed for 40 hrs.



Note:

Must use FRESH TsCl. Total yield 150 mg.

50/50 Et₂O/H₂O



200 mg of Tris- $(\text{CH}_2)_6\text{OH}$ (2.13 mols)

276.40

- 1.01

275.39 g

516.89

3 ml

10 mM TEAE

10 mM AgNO_3

140 mM DTT

1 mg dissolved in 1 ml (5 ml cold, adding TEAE buffer)

1 ml of AgNO_3 solution - 30 min

1 ml of DTT - 30 min (54.2 mM) 152.5 ml

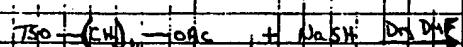
= 1M

1 ml of 100 mM DTT or 1/1 ml of 1M DTT

1 ml of 10 mM AgNO_3 or 1/1 ml of 1M AgNO_3

NMR says NR. The majority of isolated material is strongly nitrated

144



1 equiv

2 equiv

0.

NaI + Anisole
(below)

$\mu = 454.72$

or 3.3×10^{-4}
or 18.4 cm^{-1}

NMR reveals that the
spot (circled) is from
the reaction of triethyl OH
with AcOH did not occur

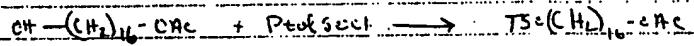
NaI = triethyl OH NMR

SD:ED
Dilution : 1:40000

$\approx 75 \text{ mgs}$ starting material $\approx 1.7 \times 10^{-4} \text{ mols} \times 1.2 \text{ equiv of NaBH}_4$

2×10^{-4} mols NaBH₄ or 8 mgs dissolved in MeOH

50:50
AcOH / H₂O



2 x 250 mgs or 8.5×10^{-3} moles in 15 ml of dry pyridine is cooled to 0°C. An ice bath. 325 mgs of TSOCl is added and the reaction allowed to proceed @ 4°C. The product (brownish tint) is poured into a beaker with 100 ml of ice water, stirred for 10 min and filtered. The solid is washed with water and dissolved in pet ether and charcoal added with stirring. The mixture was filtered and dried (40 mgs total).

~~2.5 hours is not long enough, see one week~~



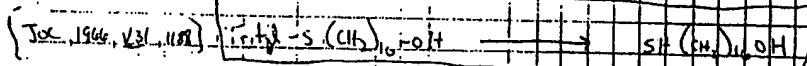
325 mgs ($\text{m.w.} = 454.72$) or 7.15×10^{-3} moles was dissolved in 10 ml of dry DMF and degassed. Triethyl methyl boronate ($\text{m.w.} 276.4$) with 1.1 equiv (7.86×10^{-3}) or 217 mgs and 1.05 equiv of NaOH or 30 mgs was dissolved in 150 ml of H_2O and degassed.

The triethyl SH was dissolved in 5 ml of EtOH and degassed. The NaOH was added ^{1/2} at a time to the EtOH solution. Then the $\text{TSO}-\text{COCl}$ added. The reaction was rapidly degassed and allowed to proceed for 12 hrs.

The reaction was TiCl_4 and NaOH added to precipitate.

Flask \rightarrow phosphonamide

EtoAc



MW = 516.89

25 mgs (4.16 $\times 10^{-3}$ mols) ^{LiCl} was dissolved in ^{1 ml} acetic acid (1.15 ml) and .39 mls of 1N HCl added. The reaction was allowed to stir for 15 hrs and 75°C . Upon addition of 1N, I ¹ ml was found a white ^{solid} precipitate. After stirring for 5 hrs the ppt. redissolved with the resulting solution slightly cloudy.

Tos - 64%₁₀ C.H.C. $\xrightarrow{\text{Dry DMF}}$

NaOH

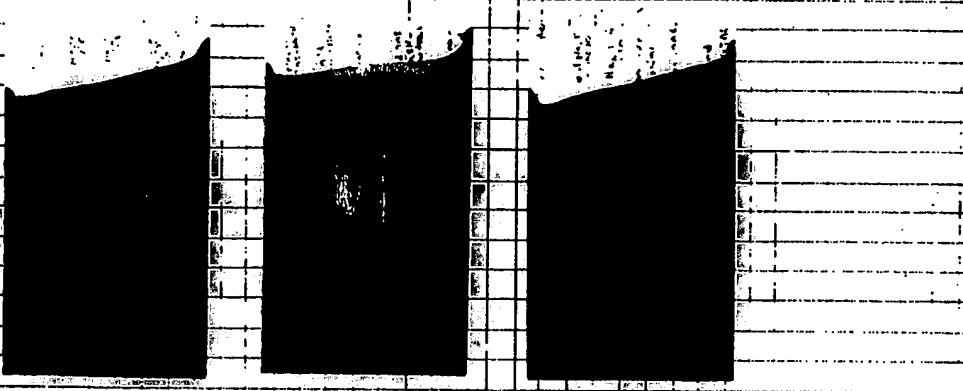
M.W. = 167.72

On 65 mg of the Tos derivative was dissolved in 5 ml of dry DMF.

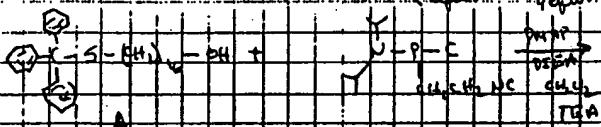
1.4×10^{-4} mols $\times 2$ equiv of NaOH = 2.9×10^{-4} mols or 17 mg.

17 mg of NaOH is dissolved in 1 ml of dry NaOH. The NaOH (in 2 ml of dry NaOH) was dispensed and the NaOH added.

This solution is added (with syringe) to the dry DMF Tos product.



148.



170 mg of A = 3.4 g/mol

151 ml of 2 equiv of phosphorous

240 ml 10% DIET

170 mg of A was dissolved in ~~200 ml~~ 45 ml of CH_2Cl_2 and

240 ml of DIET added. 151 ml of phosphorous were added

drop wise over 30 min.

The column was run with

90:10:0.5

10% DIET: 10% TEA

200 ml of wash

product

0 0

50:50:0

50:50:1%

TEA

0 0

isolated

$\approx 125 \text{ mg}$

Monoglyceride

solid: 16 mm

triglyceride

0

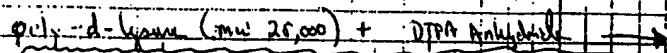
- A plate carrying Carboxymytoxin Nitrate recorded on the presence of DTT

After cooling a white ppt. formed all the solution was dissolved in 100 ml's of CH_3CO_2 pH NaCO_3 (5M, pH 9) was used to wash with CH_3CO_2 . The solution was centrifuged until the water layer was pH 8. The solution was then washed with pH 7 buffer, returned to dryness

→ Before base wash

— after washing (centrifuged) with pH 7 NaCO_3

150.



Polylysine was dissolved in DMSO and (wspms in 2 ml) and applied to a pH 10. An additional 1/2 ml was added and 3 ml total collected from each of 3 100 to 10 columns. The recovered dried material (≈ 6.5 mg) was divided into 3 reaction vessels of 2.2 mg each.

The following ratios will be prepared: 50x

(See page)
100x
200x

0.1 mM poly-d-lysine:

2.2 mg (8.6×10^{-5} mols) in 8.6 ml = 0.1 mM

50x DTPA Anhydride or $(4.3 \times 10^{-5}) \times 357.3 = 15.1 \text{ mg}$

100x 1 or $(8.6 \times 10^{-5}) \times 357.3 = 30.1 \text{ mg}$

200x 1 or $(1.72 \times 10^{-4}) \times 357.3 = 61.5 \text{ mg}$

0.5 M HCO_3 buffer, pH ≈ 9.5

Only the 200x reaction showed any change in pH (e.g. $\rightarrow 9.4$)

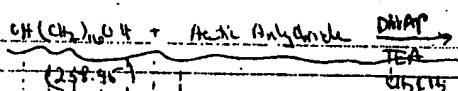
The reaction was ~~extremely~~ slow and had to dry over

400x

35.3 mg or 1.4×10^{-6} mols

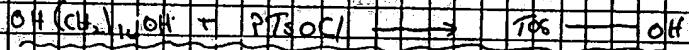
200 mg of DTPA Anhydride in 14.26 pH 9.5 molar

Buffer



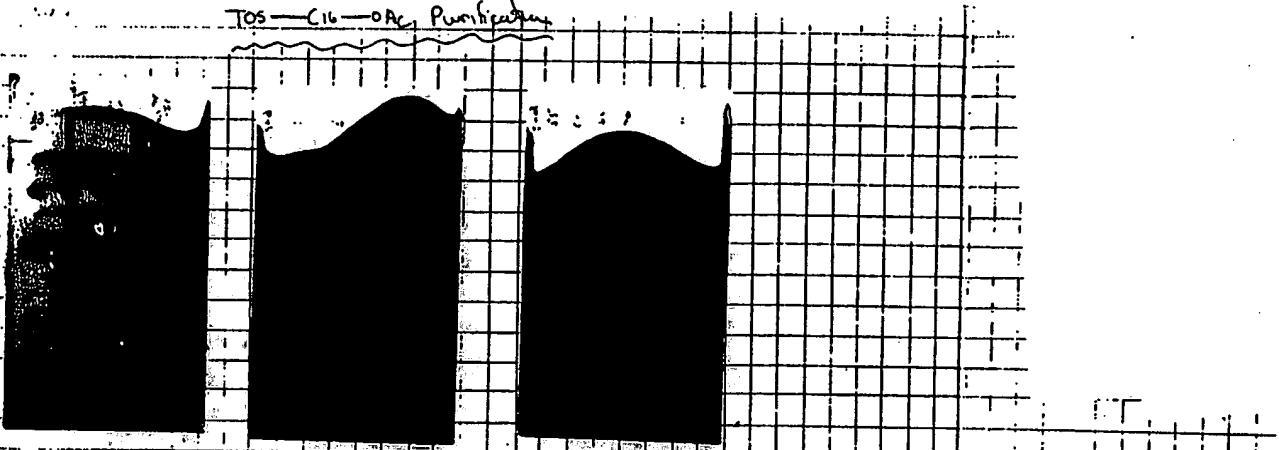
(258.95)

3 gr of $\text{CH}(\text{CH}_3)_2\text{OH}$ were dissolved in 50mls of dry CH_3Cl and
7 mg of DMAP added along with 2.5 ml of TEA
and 1.07 ml of Acetic Anhydride.



2 gr of $\text{CH}(\text{CH}_3)_2\text{OH}$ (7.74×10^{-3} mols) was dissolved in 25 ml
of dry CH_3Cl and cooled to 0°C . TsOCl (190.65) (requir
= 1.48 gms) was added and the mixture allowed to proceed
for

TOS-C16-OAc Purification



The water/egy solution is expected to dry over and stored in
hexane/ethyl ether and rehydrated 3 times. The solution is
dissolved in 50:50 DMSO:hexane and filtered. The solution is
applied to a column (cat. 5 max: Ettex 100% DMSO) several
times. Future: Run 90:10 to start and rise up to 80:20
See page 139 for prep.

Tos-C₁₆-OH + NaI $\xrightarrow{\text{heat}}$ C₁₆-OH₂ 153

180 rpm

45°

200

250

300

350

400

450

500

550

600

650

700

750

800

850

900

950

1000

1050

1100

1150

1200

1250

1300

1350

1400

1450

1500

1550

1600

1650

1700

1750

1800

1850

1900

1950

2000

2050

2100

2150

2200

2250

2300

2350

2400

2450

2500

2550

2600

2650

2700

2750

2800

2850

2900

2950

3000

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3100

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